

available at www.sciencedirect.comjournal homepage: www.elsevier.com/locate/biochempharm

New approaches to blockade of $\alpha 4$ -integrins, proven therapeutic targets in chronic inflammation

Christiane Kummer*, Mark H. Ginsberg

Department of Medicine, University of California San Diego, 9500 Gilman Drive, 0726, La Jolla, CA 92093, United States

ARTICLE INFO

Article history:

Received 6 April 2006

Accepted 12 June 2006

Keywords:

Integrins

Chronic inflammation

Therapeutic targets

Cell migration

ABSTRACT

The recruitment of leukocytes into tissue is a pivotal step in inflammation. $\alpha 4$ -Integrins are adhesion receptors on circulating leukocytes that mediate attachment to the endothelium and facilitate their migration into the inflamed tissue. This multistep process is mediated by the interaction of $\alpha 4$ -integrins with their counter receptors VCAM-1 and MadCAM-1 that are expressed on endothelial cells. $\alpha 4$ -Integrins act as both adhesive and signaling receptors. Paxillin, a signaling adaptor molecule, binds directly to the $\alpha 4$ cytoplasmic tail and its binding is important for cell migration. Blocking the adhesive functions of $\alpha 4$ -integrins has been shown to be an effective therapeutic approach in the treatment of autoimmune diseases, but also carries the risk of defects in development, hematopoiesis and immune surveillance. Interfering with $\alpha 4$ signaling by inhibiting the $\alpha 4$ –paxillin interaction decreases $\alpha 4$ -mediated cell migration and adhesion to VCAM-1 and MadCAM under shear flow. These in vitro effects are accompanied by a selective impairment of leukocyte migration into inflammatory sites when the $\alpha 4$ –paxillin interaction is blocked in vivo. Thus, blockade of $\alpha 4$ -integrin signaling may offer a novel strategy for interfering with the functions of these receptors in pathological events while sparing important physiological functions.

© 2006 Elsevier Inc. All rights reserved.

Contents

1. Introduction	1460
2. $\alpha 4$ -Integrins in adhesion and migration: function blocking antagonists	1461
3. $\alpha 4$ -Integrin signaling: an alternative approach	1462
4. Conclusions	1465
References	1465

1. Introduction

Autoimmune diseases are characterized by an inappropriate response of the immune system against self which leads to inflammation-induced dysfunction and ultimately to the destruction of the affected tissue. In rheumatoid arthritis

for example, the immune system attacks the synovium [1] while in inflammatory bowel disease (Crohn's disease) the intestinal mucosa is affected [2–4]. In multiple sclerosis T cells, B cells, macrophages and microglia mount a concerted attack against the myelin sheath surrounding the nerve fibers in the brain and spinal cord [5–8], whereas in type 1 diabetes mellitus

* Corresponding author. Tel.: +1 858 822 6496; fax: +1 858 822 6458.

E-mail address: ckummer@ucsd.edu (C. Kummer).

0006-2952/\$ – see front matter © 2006 Elsevier Inc. All rights reserved.

doi:10.1016/j.bcp.2006.06.014

destruction of the insulin producing β -cells of the pancreatic islets is mediated largely by T lymphocytes [9–11].

Although the exact causes of most chronic inflammatory diseases are unclear, leukocyte accumulation in the affected tissues or organs contributes to the pathogenesis of the disease [12,13]. The recruitment of leukocytes from the blood into the tissue at sites of inflammation is regulated by sequential engagement of adhesion and signaling molecules on leukocytes and endothelial cells such as $\alpha 4$ -integrins and their ligands [14,15]. In this review we discuss recent findings on the role of $\alpha 4$ -integrins in chronic inflammatory diseases and the impact of anti $\alpha 4$ -integrin therapy to treat these disorders. Furthermore, we describe the importance of $\alpha 4$ -integrin signaling in the immune response and thus in autoimmune diseases and highlight the possible usefulness of $\alpha 4$ -integrin signaling as a therapeutic target.

2. $\alpha 4$ -Integrins in adhesion and migration: function blocking antagonists

Integrins are cell surface receptors, which mediate cell adhesion and migration and regulate cell growth and survival. They are heterodimers consisting of α and β subunits. Each subunit contains an extracellular domain involved in ligand binding, a single transmembrane domain, and a cytoplasmic domain, which regulates integrin function. Integrins function as bi-directional signaling molecules [12,16], and binding to their ligands results in intracellular signals and conversely, cellular signaling events can modulate the affinity of integrins for extracellular ligands.

The $\alpha 4$ -integrins, $\alpha 4\beta 1$ (very late antigen-4: VLA-4) and $\alpha 4\beta 7$ are most prominent on mononuclear leukocytes, but can also be expressed on neutrophils [17]. $\alpha 4\beta 1$ mediates cells adhesion to vascular cell adhesion molecule-1 (VCAM-1) and to an alternatively spliced form of the extracellular matrix protein fibronectin (FN) [18–21]. $\alpha 4\beta 7$ is important in lymphocyte homing to mucosal tissue by adhering to the gut homing receptor mucosa addressin cell adhesion molecule (MadCAM) [22] and it also binds to VCAM-1 and FN [23,24].

$\alpha 4$ -Integrins are essential for embryogenesis, hematopoiesis, lymphocyte homing and the recruitment of leukocytes to sites of inflammation [14,21]. $\alpha 4$ -Integrins are involved in the pathogenesis of chronic inflammatory diseases such as rheumatoid arthritis [25], diabetes type 1 [26,27], inflammatory bowel disease [28–31] and multiple sclerosis. Migration of circulating leukocytes from the blood into sites of inflammation is a multistep event that involves sequential leukocyte–endothelial interactions. This process includes initial tethering to and rolling along the vascular endothelial surface, leukocyte stimulation, primarily of integrin molecules, firm adhesion and spreading to the endothelium, and finally migration across the endothelium (diapedesis) [21,32–34]. $\alpha 4$ -Integrins and their endothelial counter receptors have a unique role in this multistep cascade because they are the only molecules known to mediate both rolling (when the integrins are in a low-affinity state) and arrest (when they are in a high-affinity state) [35,36].

Because of the important role of $\alpha 4$ -integrins in cell trafficking during inflammatory processes and in autoimmune diseases they may be useful as targets in the treatment

of these disorders. A multitude of papers have been published during the last decade describing monoclonal antibodies or small molecules that are directed against $\alpha 4$ -integrins or their endothelial ligands (reviewed by [37,38]).

Monoclonal antibodies directed against $\alpha 4$ -integrins or their cell adhesion molecule ligands have been shown to be effective modulators in animal models for autoimmune diseases such as asthma [39], rheumatoid arthritis [40–43], inflammatory bowel disease [44] or diabetes type 1 [45].

Early studies provided first evidence for the importance of anti- $\alpha 4$ antibodies in inflammatory bowel disease. Administration of HP1/2, an anti- $\alpha 4$ antibody that inhibits $\alpha 4$ binding to VCAM-1 and FN [46] to the cotton-top Tamarin, a primate that develops spontaneous acute and chronic colitis that resembles ulcerative colitis, resulted in significant attenuation of inflammation [31]. Further studies on the effect of antibodies against $\alpha 4$ -integrin, $\alpha 4\beta 7$ integrin or its ligand MadCAM-1 in animal models of colitis report reduced T cell mediated intestinal inflammation and abrogated symptoms of colitis [28–31]. The promising result of these pre-clinical studies in animals led to human trials using humanized $\alpha 4$ -integrin antibodies. LDP-02 (MLN02), a $\alpha 4\beta 7$ blocking antibody was successfully used in a phase I/II clinical trial of ulcerative colitis and Crohn's disease [47,48].

Efficacy of antibodies directed against $\alpha 4$ -integrin was also studied in models for autoimmune encephalomyelitis (EAE), an autoimmune disorder with similarities to multiple sclerosis. In Lewis rats, the antibody HP1/2 completely prevented the development of paralysis in 75% of the treated animals. In those that developed the disease, paralysis was delayed and its severity was reduced [49]. Another antibody, AN100226m, was studied in a guinea pig model of EAE and administration prevented leukocyte infiltration and suppressed clinical and pathological features of EAE [50]. The humanized version of this antibody, natalizumab [51–53], has been successful in the treatment of multiple sclerosis and Crohn's disease in humans [54,55]. However, the mechanisms by which natalizumab exerts its beneficial effects are poorly understood. It is known though that it binds to $\alpha 4$ and thereby inhibits the interaction between $\alpha 4\beta 1$ and VCAM-1 and between $\alpha 4\beta 7$ and MadCAM-1 [56–58]. It has been suggested, that by inhibiting the interaction of $\alpha 4\beta 1$ and VCAM, natalizumab blocks leukocyte trafficking across the blood–brain barrier and thereby moderates inflammation within multiple sclerosis lesions. In Crohn's disease, the major players of the inflammatory process are neutrophils. Since $\alpha 4$ -integrin expression is very low on circulating neutrophils [21], it is believed, that in Crohn's disease natalizumab functions by inhibiting T lymphocytes that induce cytokines and chemokines needed to sustain neutrophil recruitment [59].

To date, approximately 8000 patients have received natalizumab for the treatment of multiple sclerosis or Crohn's disease. However, the occurrence of three cases of progressive multifocal leucoencephalopathy (PML) and their association with anti- $\alpha 4$ therapy [60–62] led to suspension of natalizumab from sales and clinical trials in February 2005 [63,64]. Clinical trials have been revived in February 2006 and in June 2006 the FDA approved an application for resumed marketing of natalizumab with a special restricted distribution program [65]. Nevertheless, it is possible that in some patients long-term $\alpha 4$ -integrin

blockade can lead to impaired immune surveillance of the central nervous system (CNS) by inhibiting $\alpha 4$ -integrin mediated immune cell trafficking to the CNS and thus allowing JC virus replication [60–62,66].

Although clinical trials using $\alpha 4$ -integrin blocking antibodies look very promising this approach also carries limitations such as high cost, potential immunogenicity of the antibodies and the requirement for intravenous administration. Thus, small molecule antagonists of the $\alpha 4$ -integrin interaction with VCAM-1 or MadCAM appear attractive in the treatment of autoimmune disorders, since their synthesis is less expensive and they can be orally administered.

Adhesion antagonists have been designed to inhibit binding of $\alpha 4\beta 1$ to VCAM-1 (reviewed by [37]) and include peptide inhibitors and VCAM-1 binding site mimics.

Peptides based on the amino acid sequences found at protein–protein interaction sites make excellent leads for antagonist development. Early reports of $\alpha 4$ -integrin peptide antagonists were published about a decade ago and were based on an alternatively spliced connecting segment of fibronectin (CS-1) that acts as a ligand for $\alpha 4\beta 1$ and has been proven to be involved in leukocyte adhesion and migration [21]. A series of overlapping peptides of CS-1 was synthesized and their ability to inhibit $\alpha 4\beta 1$ -dependent cell adhesion to fibronectin (FN) was measured [67]. This led to the minimal tripeptide epitope LDV which inhibited cell adhesion to fibronectin. At the same time a hexapeptide comprising the LDV peptide which inhibited $\alpha 4$ -integrin binding to FN, VCAM and MadCAM was discovered by Cytel.

The RGD motif that interacts with several integrins has also been in the focus of generating inhibiting peptides of $\alpha 4\beta 1$ [68,69]. A variety of extracellular matrix proteins, including collagen [68], fibrinogen [70], vitronectin [70,71] and fibronectin [72] contain the RGD motif. From RGD peptides as a lead, cyclic RCD peptides arose that were patented for a multitude of autoimmune indications [69].

Peptidomimetic small molecule antagonists of VLA4 based on the connecting segment 1 peptide sequence of FN have been shown to be potent blockers of integrin adhesive function in vitro and show efficacy in murine contact hypersensitivity and in the sheep allergic airways model and therefore are possible candidates for clinical intervention in human asthma [73].

A dual $\alpha 4\beta 1$ and $\alpha 4\beta 7$ small molecule antagonist (TR14035) was recently published that was orally active in a model of allergic asthma in Brown Norway rats. It decreased the number of eosinophils, mononuclear cells and neutrophils in the bronchoalveolar lavage fluid and led to marked reduction of lung inflammatory lesions [74].

Taken together, antibody or small molecules that block the ligand-binding site are a proven approach to the inhibition of $\alpha 4$ -integrin function. Although this therapeutic approach may be effective, it also carries the possibility of serious side-effects:

1. Mechanism-based toxicities consequent to blockade of $\alpha 4$ -integrin functions are likely to include defects in placenta-tion, heart development [75], and development of certain cells of the hematopoietic lineages, e.g. B cells [76,77].
2. Anti-integrin antibodies can elicit immune responses. For example, 11% of patients developed antibodies to natali-

zumab during a 6-month trial [58]. Furthermore, low molecular weight integrin antagonists can induce antigenic changes in integrins [78–81] including $\alpha 4\beta 1$ [82]. These antigenic changes may lead to drug-induced antibodies and consequent cytopenias [83–85].

3. Antibody or peptide integrin inhibitors can have unexpected agonistic [86] or trans-dominant effects [78]. These effects have been proposed to account for adverse consequences of GPIIb–IIIa antagonists [87–89]. Similarly, anti $\alpha 4$ antibodies can co-stimulate T cells [90] and such effects might account for the enhanced Th1 responses elicited by such antibodies administered during symptomatic experimental allergic encephalomyelitis [91].
4. Long-term blockade of the $\alpha 4$ binding site bears the risk of opportunistic infections including PML [63,64].

3. $\alpha 4$ -Integrin signaling: an alternative approach

Integrin $\alpha 4$ -subunits regulate cell migration, cytoskeletal organization and gene expression in a distinct manner from other integrin α -subunits. They promote cell migration and antagonize cell spreading and contractility. These biological activities are a function of the $\alpha 4$ cytoplasmic domain [92,93]. The cytoplasmic tail of $\alpha 4$ -integrin binds tightly to paxillin, a cytoplasmic adaptor protein, and paxillin binding is required for the ability of $\alpha 4$ -integrin to enhance migration [94,95]. Additional evidence comes from a mutated Jurkat T cell line that expresses $\alpha 4$ -integrin with a point mutation (Y991A), which selectively disrupts paxillin binding. In this cell line $\alpha 4\beta 1$ -dependent migration but not $\alpha 4$ -dependent static adhesion, is disrupted [96]. Another signal transduction pathway that is affected by this mutation is trans-regulation of $\alpha L\beta 2$ by $\alpha 4\beta 1$ [96]. Engagement of $\alpha 4\beta 1$ by ligands such as VCAM-1 stimulates $\alpha L\beta 2$ -dependent cell adhesion and migration which is relevant to the immune response [97–100]. This form of trans-regulation in T cells requires the binding of paxillin to the $\alpha 4$ -integrin cytoplasmic domain and is abolished by disruption of this interaction by the $\alpha 4$ (Y991A) mutation. The $\alpha 4$ -paxillin interaction mediates trans-regulation by enhancing the activation of tyrosine kinases, focal adhesion kinase (FAK) and/or proline-rich tyrosine kinase-2 (Pyk2). Disruption of the paxillin– $\alpha 4$ interaction results in much less $\alpha 4\beta 1$ mediated phosphorylation of Pyk2 and FAK, and blocked $\alpha 4\beta 1$ stimulation of $\alpha L\beta 2$ -dependent migration [96]. Furthermore, $\alpha 4\beta 1$ binding to VCAM-1 or FN stimulates c-Src phosphorylation and promotes c-Src catalytic activation in an FAK-independent manner. This linkage between $\alpha 4$ and Src does not require paxillin binding to $\alpha 4$ since $\alpha 4$ WT and $\alpha 4$ Y991A expressing FAK^{−/−} cells show equal levels of membrane ruffling and motility in haptotaxis and random motility assays [101]. Transgenic mice that are homozygous for a mutation in the $\alpha 4$ -integrin cytoplasmic tail ($\alpha 4$ Y991A) that disrupts paxillin binding are viable. Their viability and fertility stands in stark contrast to the embryonic lethal $\alpha 4$ -integrin null mice. Mice with disrupted $\alpha 4$ -paxillin interaction show defective recruitment of mononuclear leukocytes in thioglycollate-induced peritonitis (Fig. 1, republished with copyright permission by JCI). Importantly, these mice have

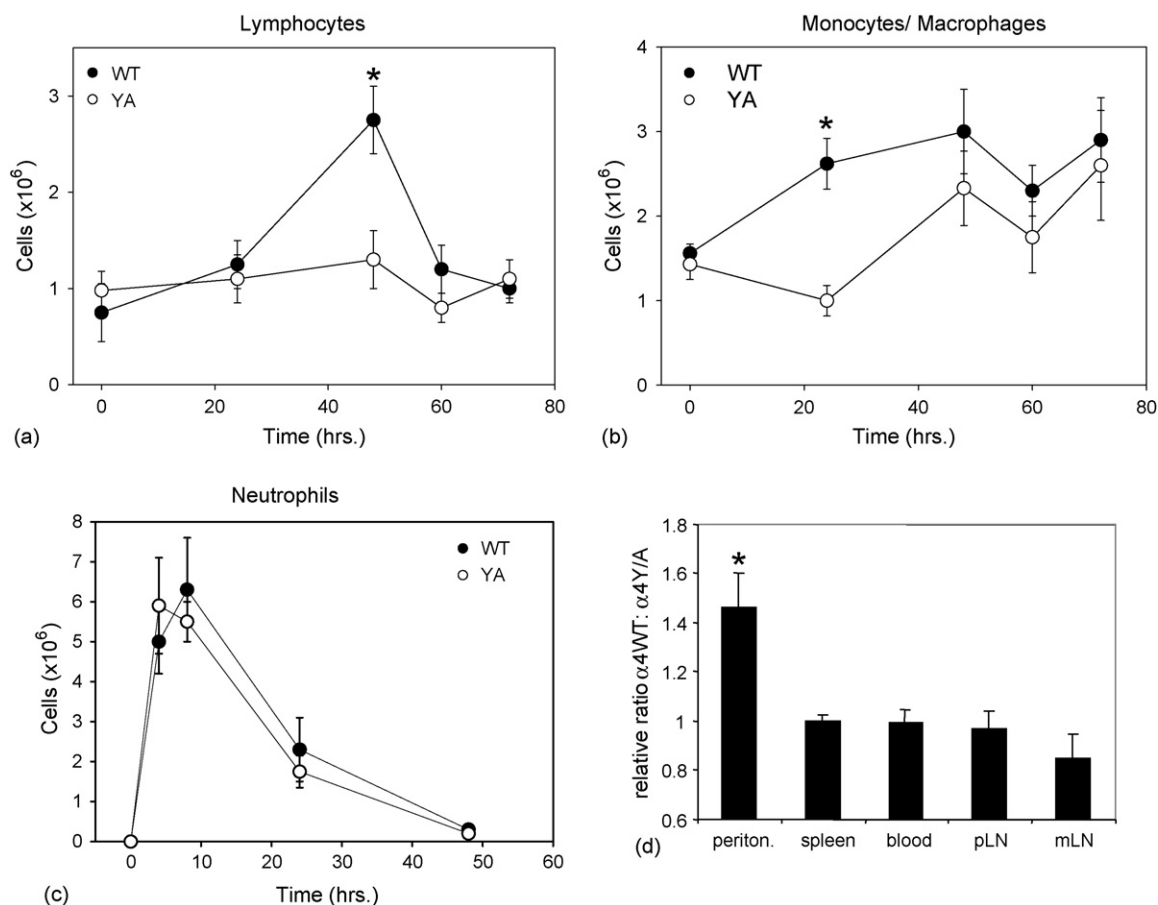


Fig. 1 – The recruitment of mononuclear leukocytes to the peritoneum in response to thioglycollate is impaired in mice with disrupted $\alpha 4$ -integrin-paxillin interaction. (a–c) WT and $\alpha 4$ (Y991A) mice were injected intraperitoneally with thioglycollate, and peritoneal lavage fluid collected at the indicated time points. Total cell number in the lavage fluid was measured with a hemocytometer, and differential cell counts were performed on cytospin slides after modified Wright–Giemsa staining. Results are shown for total lymphocyte (a), monocyte/macrophage (b), and neutrophil (c) counts. * $P = 0.013$, two-tails Student's *t*-test. Results are mean \pm S.E.M. of four to eight mice for each time point. (d) Ratios of adoptively transferred WT/ $\alpha 4$ (Y991A) splenic lymphocytes found in the spleen, blood, peripheral LN (pLN), mesenteric LN (mLN), and thioglycollate-induced inflamed peritoneal cavities (periton.) of recipient WT mice. Recipient WT mice with thioglycollate-induced peritonitis were injected with CFSE labeled splenic mononuclear cells from WT or $\alpha 4$ (Y991A) mice to test whether the defect in peritoneal lymphocytosis was ascribable to defective homing of the mutant leukocytes. Ratios of differentially labeled cells were assessed after 24 h by flow cytometry and normalized to the starting input ratio. Results are mean \pm S.E.M. of eight mice from three separate experiments. $P = 0.037$, WT vs. $\alpha 4$ (Y991A), one-tailed Student's *t*-test (figure published in Ref. [102]).

normal hemograms, normal abundance of hematopoietic precursors and unimpaired homing of hematopoietic progenitor cells to the bone marrow. These findings lend support to the idea that blockade of $\alpha 4$ -integrin signaling can impair recruitment of leukocytes to sites of inflammation while averting the adverse effects of $\alpha 4$ -integrin loss on development and hematopoiesis [102].

Paxillin is the founding member of a family of related proteins that contains two additional members, leupaxin and Hic-5 [103]. They contain N-terminal LD motifs and C-terminal LIM-domains, which mediate protein–protein interactions [104,105]. Because of the high degree of sequence similarity within the LD- and LIM-domains, it is anticipated that some of

the functions of paxillin, leupaxin and Hic-5 overlap. In addition to $\alpha 4$ -integrin, paxillin also interacts with $\alpha 9$ -integrin [94], which is most closely related to $\alpha 4$ and promotes transendothelial neutrophil migration [106]. A principal role of paxillin is to transmit signals from integrins to assure efficient cell migration.

The association of paxillin with the $\alpha 4$ tail is regulated by the phosphorylation state of Ser988 in the $\alpha 4$ tail [107]. Jurkat T cells expressing a phosphorylation-mimicking ($\alpha 4$ S998D) $\alpha 4$ variant show reduced cell migration and enhanced cell spreading due to the disruption of paxillin binding to $\alpha 4$. In contrast, Jurkat T cells that express the non-phosphorylatable $\alpha 4$ S998A mutant have a spreading defect. Surprisingly, this

mutation led to an unexpected suppression of cell migration [108]. This can be explained by the fact that efficient $\alpha 4$ -mediated cell migration requires precise spatial control of $\alpha 4$ phosphorylation by protein kinase A (PKA) and hence of paxillin binding to the $\alpha 4$ -integrin tail. $\alpha 4$ phosphorylation by PKA only occurs at the leading edge of the migrating cell which leave $\alpha 4$ -integrin unbound to paxillin resulting in Rac activity and the formation of stable lamellipodia. This spatial regulation of the $\alpha 4$ -paxillin interaction contributes to suppression of lamellipodia at the sides and rear but not at the leading edge of migrating cells and thus to more efficient cell migration [109].

It is clear now that binding of paxillin to the $\alpha 4$ tail inhibits adhesion-dependent lamellipodium formation by blocking Rac activation and the region responsible for reduction in Rac activation has been mapped to the LD4 domain of paxillin. Furthermore, Arf-GAP has been shown to be responsible for inhibition of Rac activation by decreasing Arf activity. The

localized formation of the $\alpha 4$ -paxillin-Arf-GAP complex mediates the polarization of Rac activity and directional migration [110].

These data suggest that integrin signaling might be a favorable therapeutic target for the treatment of chronic inflammatory disorders and that multiple potential targets exist within these signaling pathways. Intracellular integrin regulatory pathways are cell type-specific [16] and may therefore contain targets for cell type-specific blockade of $\alpha 4$ function, such as the $\alpha 4$ -paxillin interaction (Fig. 2). Furthermore, blockade of integrin signaling can leave ligand binding function partially intact [96,99,111]. Consequently, only partial inhibition of integrin function may occur, even with full blockade of the target. This may provide a more beneficial therapeutic ratio because mechanism-based toxicities can be easily averted. Thus, intracellular approaches may provide new and therapeutically advantageous targets for inhibition of $\alpha 4$ -integrin in autoimmune diseases.

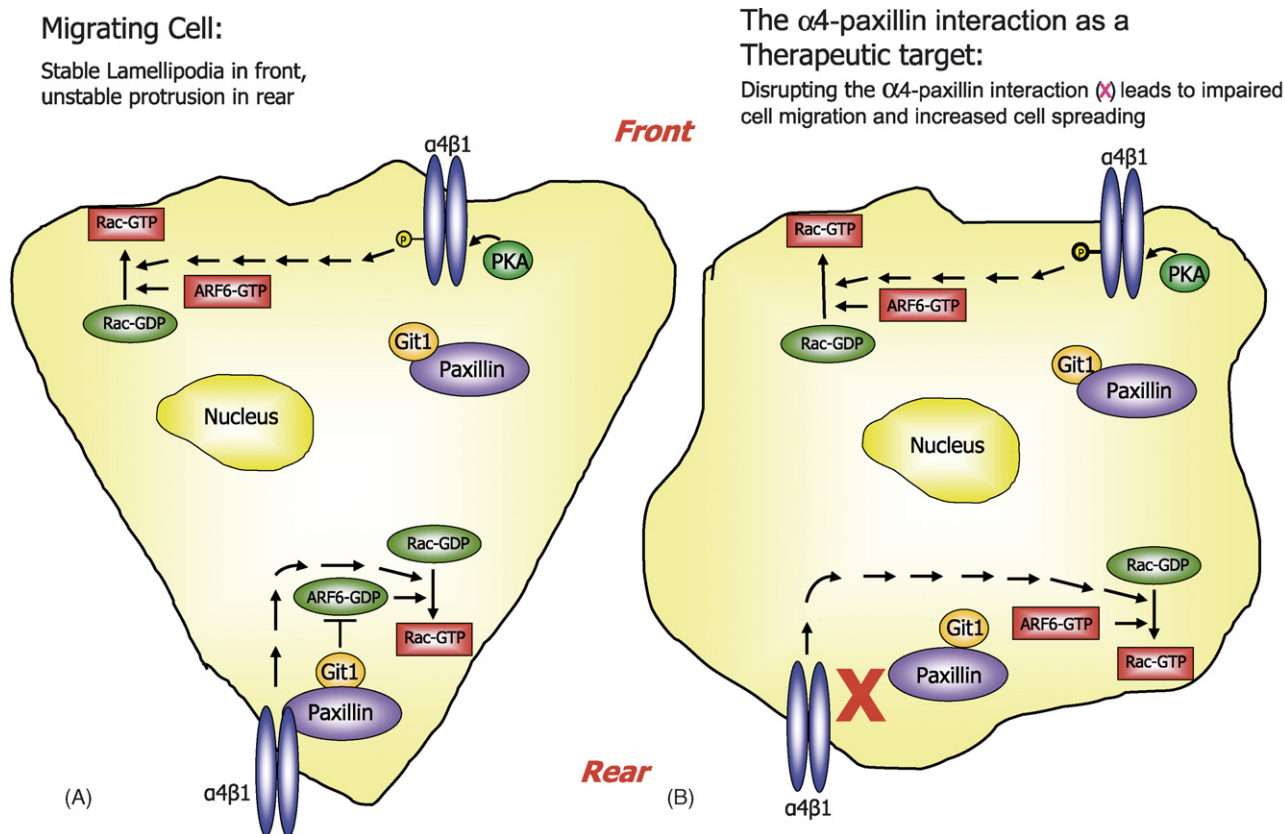


Fig. 2 – (A) Mechanism for the spatial localization of Rac activity to enhance cell migration. Formation of a stable lamellipodium at the front of migrating cells requires localization of Rac activation to the leading edge. Restriction of $\alpha 4$ -integrin phosphorylation to the leading edge limits the interaction of $\alpha 4$ with paxillin to the sides and rear of a migrating cell. The $\alpha 4$ -paxillin complex inhibits stable lamellipodia, thus confining lamellipod formation to the cell anterior. Binding of paxillin to the $\alpha 4$ -integrin subunit inhibits adhesion-dependent lamellipodium formation by blocking Rac activation. The paxillin LD4 domain mediates this reduction in Rac activity by recruiting the ADP-ribosylation factor GTPase-activating protein (Arf-GAP) Git1 that decreases Arf activity, thereby inhibiting Rac. Finally, the localized formation of the $\alpha 4$ -paxillin-Arf-GAP complex mediates the polarization of Rac activity and promotes directional cell migration. **(B) $\alpha 4$ -integrin signaling as a therapeutic target.** A mutation in the cytoplasmic tail of $\alpha 4$ -integrin (Y991A) leads to disruption of the $\alpha 4$ -paxillin interaction and thereby to reversion of inhibition of Rac activity at the rear and lateral edges of the cell. This results in decreased cell migration and enhanced cell spreading. Small molecule inhibitors that specifically target the $\alpha 4$ -paxillin interaction might have the same effect and are therefore very promising for therapeutic intervention. X represents the site of possible therapeutic intervention.

The first small molecule that inhibits the α 4-integrin-paxillin interaction has been identified by screening a positional scanning library and was published in 2002. It efficiently blocks the interaction of α 4-integrin with paxillin in vitro and impaired α 4-integrin mediated leukocyte migration [112]. Thus, small molecule interference with the α 4-paxillin interaction is feasible.

Downstream of integrins, paxillin can bind several signaling effectors to mediate migration. These effectors include the non-receptor kinase FAK [103,113,114] and its Pyk2, protein-phosphatase-PEST (PTP-PEST) [115,116] and Git1/paxillin kinase linker (PKL) [117]. FAK which is widely expressed, is recruited to focal adhesions by paxillin and adhesion stimulates FAK tyrosine phosphorylation, which can then promote cell migration [118–121]. The localization of FAK to focal adhesions seems to play a key role in FAK signaling [122] and is mediated by the focal adhesion targeting domain (FAT) of FAK [123,124]. A solution structure of the FAT domain has been described, showing that the α -helical LD2 motif of paxillin binds to the FAT domain through charge-charge and hydrophobic interactions, determining the chief residues of the LD2 peptide, involved in the interaction as R147, L148, E151, and L152 [125]. Thereby LD2 binds to α -helices H1 and H4 of FAT. This structural information is a valuable starting point to synthesize specific small molecule inhibitors that mimic the α -helical LD2 motif of paxillin and thereby interfere with FAK binding.

4. Conclusions

Blocking α 4-integrin functions has been proven useful to decrease the severity or delay the development of chronic inflammatory disorders in animal models of disease and in humans. Nevertheless, the promise of integrin-directed therapeutics has been limited by mechanism-based toxicities and even led to fatal results in a few cases. Recent findings suggest that α 4-integrin signaling might be a promising target for the treatment of autoimmune diseases. Interfering with downstream, intracellular protein-protein interactions for example that are involved in α 4-mediated cell migration raises the possibility of inhibiting cell migration but leaving α 4-mediated cell adhesion intact, and thereby obviating certain side-effects. Intriguing evidence comes from a transgenic mouse that expresses α 4-integrin with a mutation that disrupts paxillin binding. These mice exhibit defective recruitment of mononuclear leukocytes into thioglycollate-induced peritonitis, but show intact lymphohematopoiesis. Although the long-term effects of blocking α 4-integrins or their signaling are not known yet and need to be investigated in more detail, α 4-integrin signaling seems to be a promising target for the treatment of autoimmune diseases.

REFERENCES

- [1] Elices MJ. Curr Opin Anti-Inflammatory Immunomodulatory Invest Drugs 1999;1:14.
- [2] Bernstein CN. Expert Opin Ther Pat 2001;11:1081.
- [3] Podalski DK. N Engl J Med 1991;325:928.
- [4] Podolsky DK. Inflammatory bowel disease (1). N Engl J Med 1991;325(13):928–37.
- [5] Kieseier BC, Hartung HP. Current disease-modifying therapies in multiple sclerosis. Semin Neurol 2003;23(2):133–46.
- [6] Platten M, Steinman L. Multiple sclerosis: trapped in deadly glue. Nat Med 2005;11(3):252–3.
- [7] Steinman L. Multiple sclerosis: a two-stage disease. Nat Immunol 2001;2(9):762–4.
- [8] Zamvil SS, Steinman L. Diverse targets for intervention during inflammatory and neurodegenerative phases of multiple sclerosis. Neuron 2003;38(5):685–8.
- [9] Notkins AL, Lernmark A. Autoimmune type 1 diabetes: resolved and unresolved issues. J Clin Invest 2001;108(9):1247–52.
- [10] Tisch R, McDevitt H. Insulin-dependent diabetes mellitus. Cell 1996;85(3):291–7.
- [11] Delovitch TL, Singh B. The nonobese diabetic mouse as a model of autoimmune diabetes: immune dysregulation gets the NOD. Immunity 1997;7(6):727–38.
- [12] Hynes RO. Integrins: versatility, modulation, and signaling in cell adhesion. Cell 1992;69(1):11–25.
- [13] Sharar SR, Winn RK, Harlan JM. The adhesion cascade and anti-adhesion therapy: an overview. Springer Semin Immunopathol 1995;16(4):359–78.
- [14] Butcher EC, Picker LJ. Lymphocyte homing and homeostasis. Science 1996;272(5258):60–6.
- [15] von Andrian UH, Mackay CR. T cell function and migration. Two sides of the same coin. N Engl J Med 2000;343(14):1020–34.
- [16] Schwartz MA, Schaller MD, Ginsberg MH. Integrins: emerging paradigms of signal transduction. Annu Rev Cell Dev Biol 1995;11:549–99.
- [17] Hemler ME. VLA proteins in the integrin family: structures, functions, and their role on leukocytes. Annu Rev Immunol 1990;8:365–400.
- [18] Bevilacqua MP. Endothelial-leukocyte adhesion molecules. Annu Rev Immunol 1993;11:767–804.
- [19] Hemler ME, Elices MJ, Parker C, Takada Y. Structure of the integrin VLA-4 and its cell-cell and cell-matrix adhesion functions. Immunol Rev 1990;114:45–65.
- [20] Postigo AA, Teixido J, Sanchez-Madrid F. The alpha 4 beta 1/VCAM-1 adhesion pathway in physiology and disease. Res Immunol 1993;144(9):723–35.
- [21] Springer TA. Traffic signals for lymphocyte recirculation and leukocyte emigration: the multistep paradigm. Cell 1994;76(2):301–14.
- [22] Berlin C, Berg EL, Briskin MJ, Andrew DP, Kilshaw PJ, Holzmann B, et al. Alpha 4 beta 7 integrin mediates lymphocyte binding to the mucosal vascular addressin MAdCAM-1. Cell 1993;74(1):185–95.
- [23] Chan BM, Elices MJ, Murphy E, Hemler ME. Adhesion to vascular cell adhesion molecule 1 and fibronectin. Comparison of alpha 4 beta 1 (VLA-4) and alpha 4 beta 7 on the human B cell line JY. J Biol Chem 1992;267(12):8366–70.
- [24] Ruegg C, Postigo AA, Sikorski EE, Butcher EC, Pytela R, Erle DJ. Role of integrin alpha 4 beta 7/alpha 4 beta P in lymphocyte adherence to fibronectin and VCAM-1 and in homotypic cell clustering. J Cell Biol 1992;117(1):179–89.
- [25] Ginsberg MH, Diaz-Gonzalez F. Cell adhesion molecules and endothelial cells in arthritis. In: Koopman WJ, editor. Arthritis and allied Conditions. Baltimore: Williams & Wilkins; 1996. p. 479.
- [26] Yang XD, Karin N, Tisch R, Steinman L, McDevitt HO. Inhibition of insulinitis and prevention of diabetes in nonobese diabetic mice by blocking L-selectin and very late antigen 4 adhesion receptors. Proc Natl Acad Sci USA 1993;90(22):10494–8.

- [27] Yang XD, Michie SA, Tisch R, Karin N, Steinman L, McDevitt HO. A predominant role of integrin alpha 4 in the spontaneous development of autoimmune diabetes in nonobese diabetic mice. *Proc Natl Acad Sci USA* 1994;91(26):12604–8.
- [28] Hesterberg PE, Winsor-Hines D, Briskin MJ, Soler-Ferran D, Merrill C, Mackay CR, et al. Rapid resolution of chronic colitis in the cotton-top tamarin with an antibody to a gut-homing integrin alpha 4 beta 7. *Gastroenterology* 1996;111(5):1373–80.
- [29] Hokari R, Kato S, Matsuzaki K, Iwai A, Kawaguchi A, Nagao S, et al. Involvement of mucosal addressin cell adhesion molecule-1 (MAdCAM-1) in the pathogenesis of granulomatous colitis in rats. *Clin Exp Immunol* 2001;126(2):259–65.
- [30] Picarella D, Hurlbut P, Rottman J, Shi X, Butcher E, Ringler DJ. Monoclonal antibodies specific for beta 7 integrin and mucosal addressin cell adhesion molecule-1 (MAdCAM-1) reduce inflammation in the colon of scid mice reconstituted with CD45RBhigh CD4+ T cells. *J Immunol* 1997;158(5):2099–106.
- [31] Podolsky DK, Lobb R, King N, Benjamin CD, Pepinsky B, Sehgal P, et al. Attenuation of colitis in the cotton-top tamarin by anti-alpha 4 integrin monoclonal antibody. *J Clin Invest* 1993;92(1):372–80.
- [32] Butcher EC. Leukocyte-endothelial cell recognition: three (or more) steps to specificity and diversity. *Cell* 1991;67(6):1033–6.
- [33] Nourshargh S, Marelli-Berg FM. Transmigration through venular walls: a key regulator of leukocyte phenotype and function. *Trends Immunol* 2005;26(3):157–65.
- [34] Steeber DA, Venturi GM, Tedder TF. A new twist to the leukocyte adhesion cascade: intimate cooperation is key. *Trends Immunol* 2005;26(1):9–12.
- [35] Alon R, Kassner PD, Carr MW, Finger EB, Hemler ME, Springer TA. The integrin VLA-4 supports tethering and rolling in flow on VCAM-1. *J Cell Biol* 1995;128(6):1243–53.
- [36] Berlin C, Bargatze RF, Campbell JJ, von Andrian UH, Szabo MC, Hasslen SR, et al. Alpha 4 integrins mediate lymphocyte attachment and rolling under physiologic flow. *Cell* 1995;80(3):413–22.
- [37] Jackson DY. Alpha 4 integrin antagonists. *Curr Pharm Des* 2002;8(14):1229–53.
- [38] Yusuf-Makagiansar H, Anderson ME, Yakovleva TV, Murray JS, Siahaan TJ. Inhibition of LFA-1/ICAM-1 and VLA-4/VCAM-1 as a therapeutic approach to inflammation and autoimmune diseases. *Med Res Rev* 2002;22(2):146–67.
- [39] Laberge S, Rabb H, Issekutz TB, Martin JG. Role of VLA-4 and LFA-1 in allergen-induced airway hyperresponsiveness and lung inflammation in the rat. *Am J Respir Crit Care Med* 1995;151(3 Pt 1):822–9.
- [40] Dutta AS, Gormley JJ, Coath M, Hassall L, Hayward CF, Gellert PR, et al. Potent cyclic peptide inhibitors of VLA-4 (alpha4beta1 integrin)-mediated cell adhesion. Discovery of compounds like cyclo(MePhe-Leu-Asp-Val-D-Arg-D-Arg) (ZD7349) compatible with depot formulation. *J Pept Sci* 2000;6(8):398–412.
- [41] Issekutz AC, Nakazato S, Issekutz TB. Differential roles of VLA-4(CD49d/CD29) and LFA-1(CD11a/CD18) integrins and E- and P-selectin during developing and established active or adoptively transferred adjuvant arthritis in the rat. *Immunol Cell Biol* 2003;81(5):397–408.
- [42] Raychaudhuri A, Chou M, Weetall M, Jeng AY. Blockade of integrin VLA-4 prevents inflammation and matrix metalloproteinase expression in a murine model of accelerated collagen-induced arthritis. *Inflammation* 2003;27(2):107–13.
- [43] Seiffge D. Protective effects of monoclonal antibody to VLA-4 on leukocyte adhesion and course of disease in adjuvant arthritis in rats. *J Rheumatol* 1996;23(12):2086–91.
- [44] Powrie F, Leach MW. Genetic and spontaneous models of inflammatory bowel disease in rodents: evidence for abnormalities in mucosal immune regulation. *Ther Immunol* 1995;2(2):115–23.
- [45] Lobb RR, Hemler ME. The pathophysiologic role of alpha 4 integrins in vivo. *J Clin Invest* 1994;94(5):1722–8.
- [46] Pulido R, Elices MJ, Campanero MR, Osborn L, Schiffer S, Garcia-Pardo A, et al. Functional evidence for three distinct and independently inhibitable adhesion activities mediated by the human integrin VLA-4. Correlation with distinct alpha 4 epitopes. *J Biol Chem* 1991;266(16):10241–5.
- [47] Feagan B, McDonald J, Greenberg G. An ascending dose trial of a humanized alpha4beta7 antibody in ulcerative colitis (UC). *Gastroenterology* 2001;118:A874.
- [48] Lindenboom K, Brazier G. Millennium pharmaceuticals announces phase II data for MLN02 in Crohn's disease. Cambridge, MA: Millennium Pharmaceuticals; 2002.
- [49] Yednock TA, Cannon C, Fritz LC, Sanchez-Madrid F, Steinman L, Karin N. Prevention of experimental autoimmune encephalomyelitis by antibodies against alpha 4 beta 1 integrin. *Nature* 1992;356(6364):63–6.
- [50] Kent SJ, Karlik SJ, Cannon C, Hines DK, Yednock TA, Fritz LC, et al. A monoclonal antibody to alpha 4 integrin suppresses and reverses active experimental allergic encephalomyelitis. *J Neuroimmunol* 1995;58(1):1–10.
- [51] Egan LJ, Sandborn WJ. Advances in the treatment of Crohn's disease. *Gastroenterology* 2004;126(6):1574–81.
- [52] Rudick RA, Sandrock A. Natalizumab: alpha4-integrin antagonist selective adhesion molecule inhibitors for MS. *Expert Rev Neurother* 2004;4(4):571–80.
- [53] Sandborn WJ, Yednock TA. Novel approaches to treating inflammatory bowel disease: targeting alpha-4 integrin. *Am J Gastroenterol* 2003;98(11):2372–82.
- [54] Ghosh S. Therapeutic value of alpha-4 integrin blockade in inflammatory bowel disease: the role of natalizumab. *Expert Opin Biol Ther* 2003;3(6):995–1000.
- [55] Gordon FH, Lai CW, Hamilton MI, Allison MC, Srivastava ED, Fouweather MG, et al. A randomized placebo-controlled trial of a humanized monoclonal antibody to alpha4 integrin in active Crohn's disease. *Gastroenterology* 2001;121(2):268–74.
- [56] Natalizumab: AN 100226, anti-4alpha integrin monoclonal antibody. *Drugs R D* 2004;5(2):102–7.
- [57] Ghosh S, Goldin E, Gordon FH, Malchow HA, Rask-Madsen J, Rutgeerts P, et al. Natalizumab for active Crohn's disease. *N Engl J Med* 2003;348(1):24–32.
- [58] Miller DH, Khan OA, Sheremata WA, Blumhardt LD, Rice GP, Libonati MA, et al. A controlled trial of natalizumab for relapsing multiple sclerosis. *N Engl J Med* 2003;348(1):15–23.
- [59] von Andrian UH, Engelhardt B. Alpha4 integrins as therapeutic targets in autoimmune disease. *N Engl J Med* 2003;348(1):68–72.
- [60] Kleinschmidt-DeMasters BK, Tyler KL. Progressive multifocal leukoencephalopathy complicating treatment with natalizumab and interferon beta-1a for multiple sclerosis. *N Engl J Med* 2005;353(4):369–74.
- [61] Langer-Gould A, Atlas SW, Green AJ, Bollen AW, Pelletier D. Progressive multifocal leukoencephalopathy in a patient treated with natalizumab. *N Engl J Med* 2005;353(4):375–81.
- [62] Van Assche G, Van Ranst M, Sciort R, Dubois B, Vermeire S, Noman M, et al. Progressive multifocal leukoencephalopathy after natalizumab therapy for Crohn's disease. *N Engl J Med* 2005;353(4):362–8.

- [63] Steinman L. Blocking adhesion molecules as therapy for multiple sclerosis: natalizumab. *Nat Rev Drug Discov* 2005;4(6):510–8.
- [64] Strupp M. The natalizumab story. *J Neurol* 2006;253(4):541–3.
- [65] FDA Natalizumab information, <http://www.fda.gov/cder/drug/infopage/natalizumab/default.htm>; 2006.
- [66] Berger JR, Korolnik IJ. Progressive multifocal leukoencephalopathy and natalizumab—unforeseen consequences. *N Engl J Med* 2005;353(4):414–6.
- [67] Komoriya A, Green LJ, Mervic M, Yamada SS, Yamada KM, Humphries MJ. The minimal essential sequence for a major cell type-specific adhesion site (CS1) within the alternatively spliced type III connecting segment domain of fibronectin is leucine-aspartic acid-valine. *J Biol Chem* 1991;266(23):15075–9.
- [68] Cardarelli PM, Yamagata S, Taguchi I, Gorcsan F, Chiang SL, Lobl T. The collagen receptor alpha 2 beta 1, from MG-63 and HT1080 cells, interacts with a cyclic RGD peptide. *J Biol Chem* 1992;267(32):23159–64.
- [69] Lobl TJ, Chiang S, Cardarelli PM. Cyclic cell adhesion modulation compounds 1998;Application # 485019;Patent # 5721210.
- [70] Handagama P, Bainton DF, Jacques Y, Conn MT, Lazarus RA, Shuman MA. Kistrin, an integrin antagonist, blocks endocytosis of fibrinogen into guinea pig megakaryocyte and platelet alpha-granules. *J Clin Invest* 1993;91(1):193–200.
- [71] Pytela R, Pierschbacher MD, Ruoslahti E. A 125/115-kDa cell surface receptor specific for vitronectin interacts with the arginine-glycine-aspartic acid adhesion sequence derived from fibronectin. *Proc Natl Acad Sci USA* 1985;82(17):5766–70.
- [72] Makarem R, Newham P, Askari JA, Green LJ, Clements J, Edwards M, et al. Competitive binding of vascular cell adhesion molecule-1 and the HepII/IIICS domain of fibronectin to the integrin alpha 4 beta 1. *J Biol Chem* 1994;269(6):4005–11.
- [73] Lobb RR, Abraham WM, Burkly LC, Gill A, Ma W, Knight JA, et al. Pathophysiologic role of alpha 4 integrins in the lung. *Ann N Y Acad Sci* 1996;796:113–23.
- [74] Cortijo J, Sanz MJ, Iranzo A, Montesinos JL, Nabah YN, Alfon J, et al. A small molecule, orally active, alpha(4)beta(1)/alpha(4)beta(7) dual antagonist reduces leukocyte infiltration and airway hyper-responsiveness in an experimental model of allergic asthma in Brown Norway rats. *Br J Pharmacol* 2006;147(6):661–70.
- [75] Hynes RO, Wagner DD. Genetic manipulation of vascular adhesion molecules in mice. *J Clin Invest* 1996;98(10):2193–5.
- [76] Arroyo AG, Yang JT, Rayburn H, Hynes RO. Differential requirements for alpha4 integrins during fetal and adult hematopoiesis. *Cell* 1996;85(7):997–1008.
- [77] Arroyo AG, Yang JT, Rayburn H, Hynes RO. Alpha4 integrins regulate the proliferation/differentiation balance of multilineage hematopoietic progenitors in vivo. *Immunity* 1999;11(5):555–66.
- [78] Diaz-Gonzalez F, Forsyth J, Steiner B, Ginsberg MH. Trans-dominant inhibition of integrin function. *Mol Biol Cell* 1996;7(12):1939–51.
- [79] Frelinger III AL, Lam SC, Plow EF, Smith MA, Loftus JC, Ginsberg MH. Occupancy of an adhesive glycoprotein receptor modulates expression of an antigenic site involved in cell adhesion. *J Biol Chem* 1988;263(25):12397–402.
- [80] Gomez M, Luque A, del Pozo MA, Hogg N, Sanchez-Madrid F, Cabanas C. Functional relevance during lymphocyte migration and cellular localization of activated beta1 integrins. *Eur J Immunol* 1997;27(1):8–16.
- [81] Yednock TA, Cannon C, Vandever C, Goldbach EG, Shaw G, Ellis DK, et al. Alpha 4 beta 1 integrin-dependent cell adhesion is regulated by a low affinity receptor pool that is conformationally responsive to ligand. *J Biol Chem* 1995;270(48):28740–5.
- [82] Chen LL, Whitty A, Lobb RR, Adams SP, Pepinsky RB. Multiple activation states of integrin alpha4beta1 detected through their different affinities for a small molecule ligand. *J Biol Chem* 1999;274(19):13167–75.
- [83] Cines DB. Glycoprotein IIb/IIIa antagonists: potential induction and detection of drug-dependent antiplatelet antibodies. *Am Heart J* 1998;135(5 Pt 2 Su):S152–9.
- [84] Jennings LK, White MM. Expression of ligand-induced binding sites on glycoprotein IIb/IIIa complexes and the effect of various inhibitors. *Am Heart J* 1998;135(5 Pt 2 Su):S179–83.
- [85] Madan M, Berkowitz SD. Understanding thrombocytopenia and antigenicity with glycoprotein IIb-IIIa inhibitors. *Am Heart J* 1999;138(4 Pt 2):317–26.
- [86] Du XP, Plow EF, Frelinger III AL, O'Toole TE, Loftus JC, Ginsberg MH. Ligands “activate” integrin alpha IIb beta 3 (platelet GPIIb-IIIa). *Cell* 1991;65(3):409–16.
- [87] Peter K, Schwarz M, Ylanne J, Kohler B, Moser M, Nordt T, et al. Induction of fibrinogen binding and platelet aggregation as a potential intrinsic property of various glycoprotein IIb/IIIa (alphaIIbbeta3) inhibitors. *Blood* 1998;92(9):3240–9.
- [88] Peter K, Straub A, Kohler B, Volkmann M, Schwarz M, Kubler W, et al. Platelet activation as a potential mechanism of GP IIb/IIIa inhibitor-induced thrombocytopenia. *Am J Cardiol* 1999;84(5):519–24.
- [89] Peter K, Schwarz M, Bode C. Activating effects of GPIIb/IIIa blockers: an intrinsic consequence of ligand-mimetic properties. *Circulation* 2002;105(21):E180–1.
- [90] Bednarczyk JL, McIntyre BW. A monoclonal antibody to VLA-4 alpha-chain (CDw49d) induces homotypic lymphocyte aggregation. *J Immunol* 1990;144(3):777–84.
- [91] Theien BE, Vanderlugt CL, Eagar TN, Nickerson-Nutter C, Nazareno R, Kuchroo VK, et al. Discordant effects of anti-VLA-4 treatment before and after onset of relapsing experimental autoimmune encephalomyelitis. *J Clin Invest* 2001;107(8):995–1006.
- [92] Chan BM, Kassner PD, Schiro JA, Byers HR, Kupper TS, Hemler ME. Distinct cellular functions mediated by different VLA integrin alpha subunit cytoplasmic domains. *Cell* 1992;68(6):1051–60.
- [93] Kassner PD, Alon R, Springer TA, Hemler ME. Specialized functional properties of the integrin alpha 4 cytoplasmic domain. *Mol Biol Cell* 1995;6(6):661–74.
- [94] Liu S, Thomas SM, Woodside DG, Rose DM, Kiessens WB, Pfaff M, et al. Binding of paxillin to alpha4 integrins modifies integrin-dependent biological responses. *Nature* 1999;402(6762):676–81.
- [95] Liu S, Ginsberg MH. Paxillin binding to a conserved sequence motif in the alpha 4 integrin cytoplasmic domain. *J Biol Chem* 2000;275(30):22736–42.
- [96] Rose DM, Liu S, Woodside DG, Han J, Schlaepfer DD, Ginsberg MH. Paxillin binding to the alpha(4) integrin subunit stimulates LFA-1 (integrin alpha(L)beta(2))-dependent T cell migration by augmenting the activation of focal adhesion kinase/proline-rich tyrosine kinase-2. *J Immunol* 2003;170(12):5912–8.
- [97] Chan JR, Hyduk SJ, Cybulsky MI. Alpha 4 beta 1 integrin/VCAM-1 interaction activates alpha L beta 2 integrin-mediated adhesion to ICAM-1 in human T cells. *J Immunol* 2000;164(2):746–53.
- [98] May AE, Neumann FJ, Schomig A, Preissner KT. VLA-4 (alpha(4)beta(1)) engagement defines a novel activation pathway for beta(2) integrin-dependent leukocyte

- adhesion involving the urokinase receptor. *Blood* 2000;96(2):506–13.
- [99] Rose DM, Grabovsky V, Alon R, Ginsberg MH. The affinity of integrin $\alpha 4 \beta 1$ governs lymphocyte migration. *J Immunol* 2001;167(5):2824–30.
- [100] van den Berg JM, Mul FP, Schippers E, Weening JJ, Roos D, Kuijpers TW. $\beta 1$ integrin activation on human neutrophils promotes $\beta 2$ integrin-mediated adhesion to fibronectin. *Eur J Immunol* 2001;31(1):276–84.
- [101] Hsia DA, Lim ST, Bernard-Trifilo JA, Mitra SK, Tanaka S, den Hertog J, et al. Integrin $\alpha 4 \beta 1$ promotes focal adhesion kinase-independent cell motility via $\alpha 4$ cytoplasmic domain-specific activation of c-Src. *Mol Cell Biol* 2005;25(21):9700–12.
- [102] Feral CC, Rose DM, Han J, Fox N, Silverman GJ, Kaushansky K, et al. Blocking the $\alpha 4$ integrin–paxillin interaction selectively impairs mononuclear leukocyte recruitment to an inflammatory site. *J Clin Invest* 2006;116(3):715–23.
- [103] Schaller MD. Paxillin: a focal adhesion-associated adaptor protein. *Oncogene* 2001;20(44):6459–72.
- [104] Brown MC, Curtis MS, Turner CE. Paxillin LD motifs may define a new family of protein recognition domains. *Nat Struct Biol* 1998;5(8):677–8.
- [105] Dawid IB, Breen JJ, Toyama R. LIM domains: multiple roles as adapters and functional modifiers in protein interactions. *Trends Genet* 1998;14(4):156–62.
- [106] Taooka Y, Chen J, Yednock T, Sheppard D. The integrin $\alpha 9 \beta 1$ mediates adhesion to activated endothelial cells and transendothelial neutrophil migration through interaction with vascular cell adhesion molecule-1. *J Cell Biol* 1999;145(2):413–20.
- [107] Han J, Liu S, Rose DM, Schlaepfer DD, McDonald H, Ginsberg MH. Phosphorylation of the integrin $\alpha 4$ cytoplasmic domain regulates paxillin binding. *J Biol Chem* 2001;276(44):40903–9.
- [108] Han J, Rose DM, Woodside DG, Goldfinger LE, Ginsberg MH. Integrin $\alpha 4 \beta 1$ -dependent T cell migration requires both phosphorylation and dephosphorylation of the $\alpha 4$ cytoplasmic domain to regulate the reversible binding of paxillin. *J Biol Chem* 2003;278(37):34845–53.
- [109] Goldfinger LE, Han J, Kiosses WB, Howe AK, Ginsberg MH. Spatial restriction of $\alpha 4$ integrin phosphorylation regulates lamellipodial stability and $\alpha 4 \beta 1$ -dependent cell migration. *J Cell Biol* 2003;162(4):731–41.
- [110] Nishiya N, Kiosses WB, Han J, Ginsberg MH. An $\alpha 4$ integrin–paxillin–Arf–GAP complex restricts Rac activation to the leading edge of migrating cells. *Nat Cell Biol* 2005;7(4):343–52.
- [111] Ylanne J, Chen Y, O'Toole TE, Loftus JC, Takada Y, Ginsberg MH. Distinct functions of integrin α and β subunit cytoplasmic domains in cell spreading and formation of focal adhesions. *J Cell Biol* 1993;122(1):223–33.
- [112] Ambroise Y, Yaspan B, Ginsberg MH, Boger DL. Inhibitors of cell migration that inhibit intracellular paxillin/ $\alpha 4$ binding. A well-documented use of positional scanning libraries. *Chem Biol* 2002;9(11):1219–26.
- [113] Hildebrand JD, Schaller MD, Parsons JT. Paxillin, a tyrosine phosphorylated focal adhesion-associated protein binds to the carboxyl terminal domain of focal adhesion kinase. *Mol Biol Cell* 1995;6(6):637–47.
- [114] Salgia R, Avraham S, Pisick E, Li JL, Raja S, Greenfield EA, et al. The related adhesion focal tyrosine kinase forms a complex with paxillin in hematopoietic cells. *J Biol Chem* 1996;271(49):31222–6.
- [115] Shen Y, Schneider G, Cloutier JF, Veillette A, Schaller MD. Direct association of protein-tyrosine phosphatase PTP-PEST with paxillin. *J Biol Chem* 1998;273(11):6474–81.
- [116] Turner CE. Paxillin interactions. *J Cell Sci* 2000;113(Pt 23):4139–40.
- [117] Zhao ZS, Manser E, Loo TH, Lim L. Coupling of PAK-interacting exchange factor PIX to GIT1 promotes focal complex disassembly. *Mol Cell Biol* 2000;20(17):6354–63.
- [118] Bellis SL, Miller JT, Turner CE. Characterization of tyrosine phosphorylation of paxillin in vitro by focal adhesion kinase. *J Biol Chem* 1995;270(29):17437–41.
- [119] Brown MC, Perrotta JA, Turner CE. Identification of LIM3 as the principal determinant of paxillin focal adhesion localization and characterization of a novel motif on paxillin directing vinculin and focal adhesion kinase binding. *J Cell Biol* 1996;135(4):1109–23.
- [120] Schaller MD, Parsons JT. pp125FAK-dependent tyrosine phosphorylation of paxillin creates a high-affinity binding site for Crk. *Mol Cell Biol* 1995;15(5):2635–45.
- [121] Tachibana K, Sato T, D'Avirro N, Morimoto C. Direct association of pp125FAK with paxillin, the focal adhesion-targeting mechanism of pp125FAK. *J Exp Med* 1995;182(4):1089–99.
- [122] Ilic D, Almeida EA, Schlaepfer DD, Dazin P, Aizawa S, Damsky CH. Extracellular matrix survival signals transduced by focal adhesion kinase suppress p53-mediated apoptosis. *J Cell Biol* 1998;143(2):547–60.
- [123] Cary LA, Guan JL. Focal adhesion kinase in integrin-mediated signaling. *Front Biosci* 1999;4:102–13.
- [124] Schlaepfer DD, Hauck CR, Sieg DJ. Signaling through focal adhesion kinase. *Prog Biophys Mol Biol* 1999;71(3–4):435–78.
- [125] Liu G, Guibao CD, Zheng J. Structural insight into the mechanisms of targeting and signaling of focal adhesion kinase. *Mol Cell Biol* 2002;22(8):2751–60.